

# Challenges and perspectives of surgical treatment of liver failure.

## Current status and last achievements in Georgia



*Ann. Ital. Chir.*, 2021 92, 6: 595-603  
pii: S0003469X21031389

Nina Inauri\*, Keti Tsomaia\*/\*\*, Avtandil Ghirdaladze\*, Zurab Chkhaidze\*, Nodar Khodeli\*/\*\*, Iliia Chanukvadze\*\*\*, Zurab Kakabadze\*\*\*, Dimitri Kordzaia\*/\*\*, Merab Kiladze\*

\**Ivane Javakhsishvili Tbilisi State University*

\*\**Aleksandre Natishvili Institute of Morphology, Tbilisi, Georgia*

\*\*\**Tbilisi State Medical University, Tbilisi, Georgia*

### Challenges and perspectives of surgical treatment of liver failure. Current status and last achievements in Georgia

*Liver transplantation is considered to be the last hope of treatment for irreversible liver failure caused by different diffuse and/or space-occupying lesions of this organ. The strict limitation of the donor organs stipulates for development of alternative approaches for the solving this problem.*

*The presented review of literature and our experience aims to discuss the modern aspects of management of different hepatic pathologies causing liver failure with the view of creation of the auxiliary, bioengineer-based functional tissues and/or organs and innovative surgical interventions allowing to conduct the operations in cases, which were up to date considered as inoperable.*

*There are highlighted the last achievements of the experimental and translational studies performed in four University research centers of Georgia, which, on the one hand, provoke the specific professional interest, and on the other hand, require the international cooperation and collaboration for further progress and advances in this field of surgery.*

KEY WORDS: Artificial liver, Bio-Artificial organs, Liver failure, Innovative surgery, Tissue engineering

### Introduction

Liver transplantation is considered to be the last hope of treatment for irreversible liver failure caused by diffuse and/or space-occupying lesions of this organ. At the same time, the acute shortage of donor organs<sup>1</sup>, as well as the complications, accompanying the rejecting reactions of the transplanted organ and/or prolonged immunosuppression<sup>2</sup>, lead to the intensive search for alternative treatment methods.

This search develops into two strategic directions:

*Bioengineering - as an organ replacement therapy*, based on creation of auxiliary bioartificial functional tissues and/or organs in vivo and/or in vitro, which can be transplanted orthotopically or heterotopically<sup>3-9</sup>.

*Innovative surgery*, allowing liver resections in cases which were previously considered as inoperable. The following techniques obtained the particular interest:

– Extensive and atypical extracorporeal hepatic resection with the further replantation of the remained part of the organ<sup>10,11</sup>; the method has successfully passed the clinical testing and currently the experience for its standardisation has been being gained;

– Associating liver partition and portal vein ligation for staged hepatectomy (ALPPS); this approach implies ligation of the portal vein branch supplying the large volume of pathological liver tissue with its in situ separation (splitting) from normal tissue to induce rapid regeneration of future liver remnant (FLR); on the second stage the “regenerated” FLR allows the safe excision of

*Pervenuto in Redazione Dicembre 2019. Accettato per la pubblicazione Marzo 2020*

*Correspondence to: Nina Inauri, Ivane Javakhsishvili, Tbilisi State University, 1 Chavchavadze Ave, Tbilisi 0179, Georgia (e-mail: ninainauri1@gmail.com)*

the large abnormal hepatic mass without development of liver failure<sup>12-15</sup>. ALPPS methodology has been developed both in humans and in various experimental models, including small rodents<sup>16-19</sup>, which is a good prerequisite for planning and development of new researches in this direction.

The presented paper aims to review some of above-mentioned approaches for the treatment of hepatic pathologies causing liver failure. In the review there are highlighted the last achievements of the experimental and translational studies performed in the Universities/Research Centers of Georgia, which, on the one side, cause the wide professional interest, and on the other side, require the extended collaboration for further development of obtained results.

## Methods

We used the Pub Med database for our search. From 2007 to 2018, approximately 150 articles were searched with the following complexes of keywords in the title and/or abstract: 1. “liver failure” and “surgical intervention” or “surgery” or “operation”; 2. “liver Bioengineering” or “bio-artificial liver” and “liver failure”. Later, we have selected articles, published in the journals indexed in WOS and having impact factor (IF) or in Scopus. The obtained database was supplemented with those papers of Georgian researchers as well as their abstracts presented at international conferences - considered to be relevant to the aim of the given review.

## CHALLENGES AND PERSPECTIVES OF LIVER FAILURE TREATMENT

Currently more than 350 million people are suffering from chronic HBV<sup>20</sup> and about 130-170 million people are infected with chronic HCV<sup>21</sup> throughout the world. Besides, a significant number of patients do not know that they have the viral hepatitis or its complications<sup>22</sup>. WHO estimates that 1.4 million people die each year due to such complications of above-mentioned pathology as acute hepatic failure, cirrhosis or liver cancer<sup>23</sup>.

The autotransplantation of bone marrow derived stem cells (BMDSC) which is widely studied in the experiments and gradually replaced into clinical practice is considered to be one of the prospective method(s) for the treatment of cirrhosis or acute-on-chronic liver failure<sup>24-30</sup>. However, in the patients with decompensated liver cirrhosis this method is ineffective and the mortality rate ranges from 48% to 67%<sup>31,32</sup>. From this point of view liver transplantation is proved to be the last hope for their survival<sup>33</sup>. The same should be told in case of Wilson's disease when severe acute liver failure (ALF) with grade I or II encephalopathy is presented<sup>34-36</sup>, as

well as in Budd-Chiari Syndrome<sup>37</sup> or chronic hepatic failure, e.g. in long-term antitubercular treatment<sup>38,39</sup>.

It is established, that the outcome of acute liver failure is lethal in the 50-90% of cases<sup>40</sup>. Liver Transplantation for Acute or Chronic Liver Failure is Based on EASL-CLIF Diagnostic Criteria. Living donor liver transplantation (LDLT) has excellent outcomes in patients with EASL-CLIF grade 1 and 2 ALF. Without transplantation, ALF patients have a very poor prognosis<sup>41</sup>. Generally, acute-on-chronic liver failure predicts adverse outcomes after orthotopic liver transplantation. Given the dismal prognosis without transplantation, the ALF patients can be transplanted with comparably good outcomes, in particular, the patients who improve under conservative therapeutic measures<sup>42</sup>.

The deficit of donor organs and lack of timeliness prevents the application of the liver transplantation as an effective method of treatment<sup>43,44</sup>.

In spite of the fact that many countries have established the deceased donor programs, it is the remarkable deficit of transplantable livers, that continues to make the thousands of patients hopeless<sup>45</sup>.

In the developing countries, where the deceased donor programs are not established, only the half liver transplantation from living donors is remained as a real approach. The same situation is in Georgia where half liver transplantation has been performed only 31 times during 2014 – 2017 years<sup>46</sup>. All the above mentioned stipulate the attempts for seeking and development of alternative approaches for the treatment of liver failure.

## BIOENGINEERING-BASED INTERVENTIONS

The latest research confirms that a “liver bud” derived from human endogenous epithelial, endothelial and mesenchymal stem cells in vitro in three-layer culture (iPSC-LBs) can be successfully vascularized after its in vivo transplantation. The functional and structural development of the transplanted bud is similar to the liver development in embryogenesis<sup>47</sup>: in 48 hours the vascular network of the transplant becomes connected to the blood vessels of the host body and begins functioning and stimulating the further maturation of the transplanted liver bud. It begins synthesizing the liver-specific proteins - alpha-feto protein, retinol-tightening protein 4, transitate and albumin<sup>48</sup>. The similar results confirming the metabolic activity of Scaffold-free 3D bio-printed human liver tissue were obtained by Kizawa H et al. (2017)<sup>49</sup>. These results raise the hope that growth of the bio-artificial liver tissue in the 3D bio-matrixes (bio-scaffolds), including the bio-degradative ones, should be even more effective<sup>50,51</sup>.

The transplantation of in-vitro recellularized liver matrix as well as in vivo recellularization of the decellulaized single liver lobe have been studied through the experiments<sup>52-54</sup>. The results of the investigations of such

chimerical livers are quite promising, as their morphological and biochemical features reveal similarities to the native liver<sup>55</sup>. It appeared that histologically it is possible to identify the vascular and biliary structures as well as labyrinth of liver cells. In addition, the cells of these chimerical bio-artificial livers produce hepatocyte proteins and bile components and express the cytokeratins of progenitor cells and/or hepatocytes<sup>56</sup>. The functional activity of the in-vivo recellularized liver lobule was observed during 6 hours<sup>57</sup>. This finding provides the new stimuli for further studying of long-term functioning of recellularized liver or its single lobe.

It is considered that liver cells optimally realize their synthetic and metabolic functions in a typical environment<sup>58</sup>. In this regard, interesting results are promised from the repopulation of the fully decellularized human hepatic scaffold by liver stellate cells, as well as hepatocellular and hepatoblastic cancer cells, which are distinguished by high viability and proliferation. The experiments showed that the cells successfully managed not only to maintain their viability, but also started the remodeling of the matrix. This observation represents a real breakthrough on the path towards development of bioartificial liver<sup>55,59-61</sup>.

Special interest is raised by decellularized scaffolds that are obtained from non-autologous entire organs<sup>62</sup> or is created by means of artificial printing with the analogy of native liver matrix. The effectiveness of repopulation of cells in such scaffolds is largely determined by the ability of adequate vascularization<sup>63</sup>. This, in turn depends on how much the "printed" scaffold structure repeats the structure of the matrix of the native organ<sup>64</sup> and/or to which level it can activate (ensure) the membrane potential of the cells embedded in it<sup>65</sup>.

Today, the scaffolds printed with the various polymers mainly focus on the peculiarities of microcirculation, particularly, the distance between the beds of the vascular structures should not exceed 1 mm. Nevertheless the preference is given to the receipt of the entire liver decellularized scaffolds, in which not only the matrix of microcirculatory network is preserved but also the large vessels (liver artery, portal and liver veins) which could be recellularized (endothelized) by the application of endotelicocytes of the umbilical vein. It should be noted that the complete successful decellularization of liver matrix was carried out with controlled machine perfusion during several days, with increasing pressure (the outflow of 350 ml the injection solution)<sup>66</sup>.

Such perfusion can be achieved in the conditions of high-tensile strength of the above-mentioned vascular beds, which in addition to the firmness of their connective tissue structures should be also determined by sites of merging of connective tissue sheaths of portal triads and hepatic veins (intrahepatic porta-caval fibrous connections - IHPCFC) described by us previously<sup>67,68</sup> (the study was performed at Tbilisi State Medical University). It was shown IHPCFC provides especial

strength and firmness to liver matrix stipulates the remaining of wholeness of the organ in condition of different impact; Later we found IHPCFC in dogs and rats that enabled us to introduce the hypothesis about the universality of these IHPCFC in all mammals and suppose the importance of taking into account their existence while printing of liver scaffolds<sup>69</sup> (the study was performed at Institute of Morphology, Tbilisi State University).

The combining of tissue bioengineering with surgical approaches showed the effectiveness of the pancreatic islets concluded into biocontainers for treatment the type 1 diabetes. The isolated and demucosated segments of the small intestine with maintained circulation<sup>70</sup>, as well as auto vein segments were used as the biocontainers<sup>71</sup>. In the experiments conducted at Tbilisi State Medical University and at Uppsala University, pancreatic islets transplantation into the biocontainers have successfully replaced the pancreas transplantation for the treatment of insulin deficiency in case of type 1 diabetes. The same approaches were realized for the auxiliary liver<sup>72</sup>. The research hypothesis that the demucosated segment of small intestine with intact blood supply could host the transplanted liver micro fragments was tested in mice and rats<sup>71,73,74</sup>. The results of the studies (performed at Tbilisi State Medical University) confirmed that the liver tissue transplanted into intestinal segment (so call "sausage") was remodeled with the formation of new blood vessels and expression of different genes needed for angiogenesis. Transplantation of liver fragments into isolated segment of small intestine of genetically determined Dpp4-deficient rats and Non Albumin Rat (NAR) showed satisfactory metabolism, ability of synthesis and exocrine functioning, including the release of liver proteins. Similar results were obtained by Hata T, et al.<sup>75</sup>. The experimental studies showed that transplantation of liver mass (artificial liver) in the amount of 20-30% of host liver mass ensures the survival of the animals. So, it was concluded that, the liver generated in the intestinal biocontainer may be an effective alternative to the treatment of the liver failure and play the role of a "bridge" (temporary rescuer) between the therapy and transplantation.

Creation of applicable organs for transplantation by the means of scaffolds represents one of the most urgent problem(s) of modern transplantology<sup>76</sup>. The decellularized tissues of different organs were proposed for this purpose, however, finding the healthy donor tissue remains very serious problem<sup>49,60</sup>.

Taking the above mentioned into the consideration, Choi et al. proposed the creation of the model of "auxiliary liver" from the human placenta<sup>77</sup>. Some authors believe that the use of human placenta as a scaffold is the best way in tissue engineering for the creation of a new bioengineered organ, since the placenta is rich in well-developed arterial and venous vascular network, it is volumetric for seeding the transplanting cells and tis-

sues, its extracellular matrix contains numerous cytokines, chemokines and growth factors, chemotactic triggers (for instance, GCP, SDF-1) as well as triggers for angiogenesis and vasculogenesis (i.e. VEGF, HGF, EGF, FGF, PDGF, TGF-beta) <sup>73,78</sup>. It should be noted that some of these factors (VEGF, HGF, FGF, EGF, IGF-1, IGF-BP and etc.) are also featured by the hepatotrophic effect. In addition, as far as the placenta is the “organ that is being thrown away”, it is easily accessible.

Z. Kakabadze and co-authors <sup>79</sup> suggested the implantation of the liver fragments (instead of cells) in the placental scaffold, as far as these fragments contain all types of liver cells. The investigators transplanted in the sheep the “hepatized placenta” prepared in accordance with this principle. The decellularized placenta with implanted autologous multiple liver fragments of 1-2 mm<sup>3</sup> size was introduced subcutaneously, in the ilio-inguinal areas of 7 Turkish female sheep, weighing 15-20 kg, that have developed acute liver failure due to the previously performed 85% hepatectomy. Seven female sheep of the same weight, which had also undergone 85% partial hepatectomy, were used as a control group. All seven animals of the control group died due to the liver failure within 3 days following the surgery, while 5 animals in the experimental group (71,4%) survived (two animals died due to vascular thrombosis and transplantant necrosis). Moreover, if a healthy sheep liver weighed 877 ± 216 g (n = 3), in the animals of control group (at the time of their death) the liver weighed 114 ± 55 g (n = 7), while in “hepatized placenta” transplanted animals, at the moment of their withdrawal from the experiment (after 20 days from the surgery) the liver weighed 370 ± 78 g (n = 5); Thus, the “hepatized placenta” implanted in the animals assist the survival of the animals following the massive hepatectomies as well as the initiation of the regeneration of remained liver and may be propose as the “auxiliary liver” in treatment of ALF (experiments were conducted at Tbilisi State Medical University) <sup>79</sup>. The authors conclude that the human decellularized placenta has a suitable anatomic structure for creating the different bioengineered organs: its mechanically strong beds of blood vessels provide a good perfusion of tissues. Histological, X-ray (with contrast medium), radioactive (with the technetium (99mTc) mebrofenin and radiological (dopplerography and CT) studies have shown that the blood flow is satisfactory preserved after hepatization of the placental cotyledons.

Taking into the consideration that the in-growth of the implanted tissue into the extracellular matrix of the placental scaffold is accompanied by the migration of native endothelial and stromal cells promoting the revascularization of transplanted liver fragments, the authors suppose that the factors necessary for cells migration are maintained in the extracellular matrix of placenta <sup>77</sup>. Generally after transplantation of recellularized organs the thrombosis is developed quite often due to aggregation

of the thrombocytes in the blood vessels. This makes the additional restriction for the application of the decellularized organs for the transplantation, since limits the viability of the implanted tissue by several hours or days <sup>78</sup>. However, as far as placenta is characterized by higher mechanical resistance of the blood vessels and “binding ability” of arterial and vein defects, its transplantation should be accompanied by less complications due to the circulatory disorders <sup>80</sup>.

### Innovative surgical approaches

In the 70s of the last century, the hypotheses on the effectiveness of temporary transplantation of auxiliary liver from the deceased newborns to adult recipients in the cases of the ALF was developed.

The methods of transplantation of the still borns’ livers to adults intraperitoneally (under the own liver, for long-term support) or extraperitoneally (in the right ileac fossa, for temporary support) were developed on the cadavers and later implemented in dogs (the study was conducted in Tbilisi Institute of Surgery) <sup>81</sup>. In the control group ALF was modeled with formation of “side-by-side” porto-caval shunt followed by one-hour total ischemia of liver with further restoration of the blood supply to the organ (Misra and Diaz model) <sup>82-84</sup>. After the implementation of the described model in adult dogs, the heterotopic transplantation of the livers obtained from the puppies aged 6-60 days was performed.

All animals of the control group died in 18,5 +/-0.95 hours after the surgery because of the hepatic coma. Morphologically, the massive necrosis of liver tissue was revealed. However, some hepatocytes remained viable (potentially reversible lesion).

The animals, which underwent auxiliary liver transplantation showed the perfect bile excretion and the rapid growth of the transplant, which was significantly higher than in the normal puppies of the same age. This indicated that the newborn’s liver that was transplanted into the adult recipients went through the full functional and structural adaptation to the increased load. The hepatocintigraphy using Bengali pink I-131 has shown the normal functioning of the transplant in 48 hours after surgery and the maximal restoration of the recipient’s liver in 3-5 days following the surgery. The normalization of the most physiological and biochemical indicators of the blood were detected. The morphological study confirmed a significant restoration of the liver’s microstructure. Eight animals lived for 3-7 days after the auxiliary liver transplantation; 1 recipient who had undergone transplantectomy on the third days after transplantation, lived for seven months. It was concluded that the functional capability of liver of the newborns is sufficient for restoration of the liver functioning and maintain the homeostasis of adult recipients in case of ALF caused by severe but still potentially reversible lesion <sup>85,86</sup>.

The advanced and atypical extracorporeal liver resections with the further replantation of the organ is more and more attractive for the surgeons because the method allows the resection of advanced tumors, that were previously considered to be inoperable and the only method of their treatment was the total hepatectomy and orthotopic liver transplantation<sup>10,11</sup>.

The universal "Machine for Artificial Blood Circulation" (Geopatent # U 1888) constructed by the scientists of the Alexandre Natishvili Institute of Morphology, Ivane Javakhishvili Tbilisi State University should facilitate the further development of this elite surgical method and confirmation of its efficiency. The pump of this machine is fundamentally different from the pumps, existing on the market. The device provides perfusion with the non-pulsative (laminar) as well as pulsatile blood flow. The universality of the device is determined by the fact that it is equipped with the pulsator that can work synchronously with the heart. The device is designated for the full heart-lung bypass, for ensuring continuous in-vivo and/or ex-vivo blood supply of the organs of brain- or heart dead donors.

Such isolated perfused ex vivo liver models have been used to study drug toxicity, liver failure, organ transplantation and hepatic ablation and combine advantages of both previous models<sup>87</sup>, for the perfusional preservation of isolated organs and also for regional perfusion in the body of recipient<sup>43,88</sup>. However, the above-described device was successfully used also in experimental liver replantation on the various animals (rabbits, dogs, sheep).

On the one hand it allowed veno-venous bypass for the management of the ahepatic phase circuit (in order to minimize its negative effects), while on the other hand it provided continuous hemodynamic protection of transplant (replant) with native blood physiological flows throughout the whole surgery from the moment of explantation up to the end of replantation<sup>89,90</sup>, thus excluding the reperfusion lesion of the replanted organ. Its blood vessels were switched to the venous and arterial lines of the perfusion system step by step. The splanchnic blood of the recipient was supplied to the portal vein with the manageable flow while the liver artery was supplied with the blood from the caudal vena cava, additionally provided by artificial pulsating and oxygenation<sup>43,91</sup>.

Recently the interest of the scientists in this field has increased when the the same device was used for liver preservation by extracorporeal perfusion<sup>92</sup> (the study was performed at Tbilisi State University).

The investigation was performed on 6 sheep with simulated cardiac arrest and undergone 8-hours extracorporeal circulation. The device was connected to the body through the femoral artery and vein with special canulas. The biopsy of the liver was performed before the starting of perfusion, and on 4 and 8 hours of the experiment. The histological slices were stained by H&E and

were assessed by standard criteria: the degrees of steatosis (large-droplet macrovesicular steatosis [ld-MaS] and/or small-droplet macrovesicular steatosis [sd-MaS]), mononuclear portal inflammatory cell infiltration, bile ductular proliferation, cholestasis, venous congestion and hepatocellular necrosis.

Before the perfusion, no venous congestion, hepatocellular necrosis or ld-MaS were observed; less than 3% of cells were suffered by sd-MaS; mononuclear portal inflammatory cell infiltrates were found only in several areas. Mild mixed ld-MaS and sd-MaS was found in less than 5 % and 10% of the cells accordingly on the 4 and 8 hours after in vivo Machine perfusion. Similarly the mild venous congestion was present in 1 out of 6 livers after 4-hours perfusion and in 2 out of 6 livers after 8-hours Perfusion. The number of necrotic hepatocytes and portal triads infiltrated with mononuclear cells did not exceed 10% and 15% accordingly. However, there were no differences in the degree of biliary damage – cholestasis or ductular proliferation - correlating with the terms of the experiment.

Taking into the account all internationally accepted criteria of donor liver histological assessment, 8-hour in vivo perfusion of the liver in Cardiac Death Donors by using of the machine providing the pulsatile blood flow guarantees the satisfactory preservation of liver making it useful for successful transplantation.

## **Conclusion**

Bioengineering based interventions as well as innovative surgical approaches continue to be important methods alternative to-or supplemented the liver transplantation for treatment of acute/chronic liver failure.

We believe that the studies, that have been conducted at the universities/research centers of Georgia, should contribute to the determination of the future perspectives in this direction. These investigations, on the one hand, provoke the specific professional interest, and on the other hand, require the international cooperation and collaboration for further progress and advances in this field.

## **Riassunto**

Il trapianto di fegato è considerato l'ultima speranza di trattamento per l'insufficienza epatica irreversibile causata da diverse lesioni diffuse e/o occupanti spazio di questo organo. La rigorosa limitazione degli organi donatori prevede lo sviluppo di approcci alternativi per la risoluzione di questo problema.

La revisione della letteratura presentata e la nostra esperienza hanno lo scopo di discutere gli aspetti moderni della gestione di diverse patologie epatiche che causano insufficienza epatica al fine di creare tessuti e / o organi

funzionali ausiliari, basati sul bioingegnere e innovativi interventi chirurgici che consentano di condurre le operazioni in casi che erano stati considerati inoperabili. Nella rassegna vengono in particolare evidenziati i risultati degli studi sperimentali e traslazionali condotti in quattro centri di ricerca universitari della Georgia, che, da un lato, suscitano l'interesse professionale specifico e dall'altro richiedono la cooperazione e la collaborazione internazionale per ulteriori progressi e progressi in questo campo della chirurgia.

## References

- Klein S, et al.: *Organ donation and utilization in the United States, 1999-2008*. Am J Transplant, 2010; 10: 973-86, doi: 10.1111/j.1600-6143.2009.03008.x.
- Kuo TK, et al.: *Stem cell therapy for liver disease: Parameters governing the success of using bone marrow mesenchymal stem cells*. Gastroenterology, 2008; 134, 2111-2121, 2121 e2111-113, doi: 10.1053/j.gastro.2008.03.015.
- Bishop ES, et al.: *3-D bioprinting technologies in tissue engineering and regenerative medicine: Current and future trends*. Genes Dis, 2017; 4: 185-195, doi: 10.1016/j.gendis.2017.10.002.
- Camp J J P: *Development of simple 3D-printed scaffolds for liver tissue engineering*, Massachusetts Institute of Technology, 2002.
- Chen HS, et al.: *Randomized trial of spheroid reservoir bioartificial liver in porcine model of posthepatectomy liver failure*. Hepatology, 2019; 69: 329-42, doi: 10.1002/hep.30184.
- Kim Y, Ozer S, Uygun BE: In Orlando G, Lerut J, Soker S, Stratta RJ(eds): *Regenerative medicine applications in organ transplantation* (eds) Academic Press, 2014; 333-52.
- Modi, A., Verma SK, Bellare J: *Hydrophilic ZIF-8 decorated GO nanosheets improve biocompatibility and separation performance of polyethersulfone hollow fiber membranes: A potential membrane material for bioartificial liver application*. Materials science & engineering, C, Materials for biological applications, 2018; 91: 524-40, doi: 10.1016/j.msec.2018.05.051.
- Sakiyama R, et al.: *Evaluation of the mass transfer rate using computer simulation in a three-dimensional interwoven hollow fiber-type bioartificial liver*. Biotechnology letters, 2018; 40: 1567-578, doi: 10.1007/s10529-018-2609-1.
- Yan Q, et al.: *Establishment and characterization of an immortalized human hepatocyte line for the development of bioartificial liver system*. Cytotechnology, 2018; 70, 665-74, doi: 10.1007/s10616-017-0167-3.
- Mao L, et al.: *Extracorporeal hepatic resection and autotransplantation for primary gastrointestinal stromal tumor of the liver*. Transplant Proc, 2015; 47: 174-78, doi: 10.1016/j.transproceed.2014.09.111.
- Wen PH, et al.: *Extracorporeal hepatic resection and autotransplantation using temporary portocaval shunt provides an improved solution for conventionally unresectable HCC*. Dig Dis Sci, 2013; 58: 3637-640, doi: 10.1007/s10620-013-2801-z.
- Aloia TA, Vauthey JN: *Association of liver partition and portal vein ligation for staged hepatectomy (ALPPS): what is gained and what is lost?* Ann Surg, 2012; 25: e9; author reply e16-19, doi: 10.1097/SLA.0b013e318265fd3e.
- Alvarez FA, Ardiles V, Sanchez Claria, R, Pekolj, J de Santibanes E: *Associating liver partition and portal vein ligation for staged hepatectomy (ALPPS): tips and tricks*. J Gastrointest Surg, 2013; 17: 814-21, doi: 10.1007/s11605-012-2092-2.
- Clavien PA, Lillmoen KD: *Associating liver partition and portal vein ligation for staged hepatectomy*. Ann Surg, 2016; 263: 835-36, doi: 10.1097/SLA.0000000000001534.
- Wu H, Pan, G. iIn: *Operative techniques in liver resection* (ed Lunan Yan) Ch. Chapter 26, 249-54 (Springer Netherlands, 2016).
- Almau Trenard H.M., et al.: *Development of an experimental model of portal vein ligation associated with parenchymal transection (ALPPS) in rats*. Cir Esp, 2014; 92: 676-81, doi: 10.1016/j.ciresp.2013.11.005.
- Budai, A. et al.: *Animal Models for Associating Liver Partition and Portal Vein Ligation for Staged Hepatectomy (ALPPS): Achievements and Future Perspectives*. Eur Surg Res, 2017; 58: 140-57, doi: 10.1159/000453108.
- Wei W, et al.: *Establishment of a rat model: Associating liver partition with portal vein ligation for staged hepatectomy*. Surgery, 2015; 159, 1299-307, doi: 10.1016/j.surg.2015.12.005.
- Zhang GQ, Zhang Z, Lau WY, Chen Xp: *Associating liver partition and portal vein ligation for staged hepatectomy (ALPPS): A new strategy to increase resectability in liver surgery*. Int J Surg, 2014; 12: 437-41, doi: 10.1016/j.ijssu.2014.03.009.
- Ghaziani T, Sendi H, Shahraz P, Zamor P: *Hepatitis B and liver transplantation: Molecular and clinical features that influence recurrence and outcome*. World J Gastroenterol, 2014; 20: 14142-4155, doi: 10.3748/wjg.v20.i39.14142.
- Papatheodoridis G, Hatzakis A. Public health issues of hepatitis C virus infection. *Best Pract Res Clin Gastroenterol*, (2012) 26, 371-380, doi: 10.1016/j.bpg.2012.09.012.
- Kretzer IF, et al.: *Hepatitis C worldwide and in Brazil: Silent epidemic. Data on disease including incidence, transmission, prevention, and treatment*. Scientific World Journal, 2014; 2014: 827849, doi: 10.1155/2014/827849.
- Rajvanshi P, Kerr A, Bhargava, KK, Burk RD, Gupta S: *Efficacy and safety of repeated hepatocyte transplantation for significant liver repopulation in rodents*. Gastroenterology, 1996; 111: 1092-1102, doi: 10.1016/s0016-5085(96)70078-1.
- Izzy M, Watt KDL (Editorial) *Transplantation in the setting of acute-on-chronic liver failure-calculating chances*. Alimentary pharmacology & therapeutics, 2018; 48: 99-100, doi: 10.1111/apt.14679.
- Jang YO, et al.: *Effect of bone marrow-derived mesenchymal stem cells on hepatic fibrosis in a thioacetamide-induced cirrhotic rat model*. BMC Gastroenterol, 2014; 14, 198, doi: 10.1186/s12876-014-0198-6.
- Jin S Z, et al.: *Stromal cell derived factor-1 enhances bone marrow mononuclear cell migration in mice with acute liver failure*. World J Gastroenterol, 2009; 15: 2657-664, doi: 10.3748/wjg.15.2657.
- Obermajer N, Popp FC, Johnson, CL, Benseler V, Dahlke MH: *Rationale and prospects of mesenchymal stem cell therapy for liver transplantation*. Current opinion in organ transplantation, 2014; 19: 60-64, doi: 10.1097/MOT.0000000000000031.

28. Piscaglia AC, Campanale M, Gasbarrini A, Gasbarrini G.: *Stem cell-based therapies for liver diseases: State of the art and new perspectives*. Stem cells international, 2010; 2010: 259461, doi: 10.4061/2010/259461.
29. Volarevic V, Nurkovic J., Arsenijevic N, Stojkovic M.: *Concise review: Therapeutic potential of mesenchymal stem cells for the treatment of acute liver failure and cirrhosis*. Stem cells, 2014; 32: 2818-823. doi: 10.1002/stem.1818.
30. Xue R, et al.: *The assessment of multipotent cell transplantation in acute-on-chronic liver failure: A systematic review and meta-analysis*. Translational research: the journal of laboratory and clinical medicine, 2018; 200;65-80, doi: 10.1016/j.trsl.2018.05.006.
31. Heise M. et al: *Liver transplantation in acute-on-chronic liver failure: considerations for a systematic approach to decision making*. Visceral medicine, 2018; 34: 291-94. doi: 10.1159/000492137.
32. Lal BB, Sood V, Khanna R, Alam, S: *How to identify the need for liver transplantation in pediatric acute-on-chronic liver failure?* Hepatology international, 2018; 12: 552-559, doi: 10.1007/s12072-018-9901-y.
33. Olivo R, Guarrera JV, Pypopoulos NT: *Liver transplantation for acute liver failure*. Clinics in liver disease, 2018; 22: 409-17. doi: 10.1016/j.cld.2018.01.014.
34. Kido J. et al.: *Recovery of severe acute liver failure without transplantation in patients with Wilson disease*. Pediatric transplantation, 2018; 22: e13292, doi: 10.1111/ptr.13292.
35. Zhang Y, et al.: *Plasmapheresis combined with continuous plasma filtration adsorption rescues severe acute liver failure in wilson's disease before liver transplantation*. Blood purification, 2019; 47: 120-25. doi: 10.1159/000493909.
36. Estrada Leon I, et al: *Urgent liver transplantation for acute liver failure due Wilson's disease*. Gastroenterologia y hepatologia, 2019; 42: 392-9. doi: 10.1016/j.gastrohep.2018.05.023.
37. Obed, A., Bashir A, Jarrad AA: *Case of live donor liver transplantation in acute-on-chronic liver failure with budd-chiari syndrome: donor and recipient with antiphospholipid antibody syndrome*. The American Journal of case reports, 2018; 19: 767-72, doi: 10.12659/AJCR.909694.
38. Li Li AA, Dibb P, Cholankeril G, Kim D, Ahmed A: *Case report of isoniazid-related acute liver failure requiring liver transplantation*. Diseases, 2018; doi: 10.3390/diseases6020040.
39. Martino RB, Abdala E, Villegas FC, D'Albuquerque, LAC, Song ajw: *Liver transplantation for acute liver failure due to antitubercular drugs. A single-center experience*. Clinics, 2018; 73: e344, doi: 10.6061/clinics/2018/e344.
40. Marudanayagam R, et al.: *Aetiology and outcome of acute liver failure*. HPB: the official journal of the International Hepato Pancreato Biliary Association, 2009; 11: 429-34. doi: 10.1111/j.1477-2574.2009.00086.x.
41. Bhatti AB et al.: *Living Donor Liver Transplantation for Acute on Chronic Liver Failure Based on EASL-CLIF Diagnostic Criteria*. Journal of clinical and experimental hepatology, 2018; 8: 136-43. doi: 10.1016/j.jceh.2017.11.007.
42. Huebener P, et al.: *Stabilisation of acute-on-chronic liver failure patients before liver transplantation predicts post-transplant survival*. Alimentary pharmacology & therapeutics, 2018; 47: 1502-510. doi: 10.1111/apt.14627.
43. Khodeli N, Chkhaidze Z., Partsakhashvili D, Pilishvili O, Kordzaia D: *Theoretical Background of Finding Organs for Transplantation among Non-Heart Beating Donors under Unsuccessful Extracorporeal Resuscitation (Literature Review)*. Georgian medical news, 2016, 92-97.
44. Matevossian, E. et al.: *Donor organ shortage crisis: a case study review of a financial incentive-based system*. Transplant Proc, (2014) 46, 2030-2035, doi: 10.1016/j.transproceed.2014.06.024.
46. *International registry in organ donation and transplantation*. 38.710 donation 102 countries reported since 1996, <http://www.irodat.org/?p=database&c=GE&year=2012#data> 2019.
47. Matsumoto K, Yoshitomi H, Rossant J, Zaret KS: *Liver organogenesis promoted by endothelial cells prior to vascular function*. Science, 2001; 294: 559-63, doi: 10.1126/science.1063889.
48. Takebe T. et al.: *Vascularized and functional human liver from an iPSC-derived organ bud transplant*. Nature, 2013; 49: 481-84; doi: 10.1038/nature12271.
49. Kizawa H, Nagao E., Shimamura M, Zhang G, Torii H: *Scaffold-free 3D bio-printed human liver tissue stably maintains metabolic functions useful for drug discovery*. Biochemistry and biophysics reports, 2017; 10, 186-91, doi: 10.1016/j.bbrep.2017.04.004.
50. Rashidi H, et al.: *3D human liver tissue from pluripotent stem cells displays stable phenotype in vitro and supports compromised liver function in vivo*. Arch Toxicol, 2018; 92: 3117-129, doi: 10.1007/s00204-018-2280-2.
51. Vishwakarma SK, et al.: *Bioengineered functional humanized livers: An emerging supportive modality to bridge the gap of organ transplantation for management of end-stage liver diseases*. World J Hepatol, 2018; 10: 822-36. doi: 10.4254/wjgh.v10.i11.822.
52. Cardoso L., Moreira LFP, Pinto MA, Henriques-Pons A, Alves: *Domino hepatocyte transplantation: A therapeutic alternative for the treatment of acute liver failure*. Canadian journal of gastroenterology & hepatology, 2018; 2593745, doi: 10.1155/2018/2593745.
53. Got'e SV, et al.: *Correction of chronic liver failure by transplantation of liver cells suspension and cell-engineering designs (experimental investigation)*. Vestnik Rossiiskoi akademii meditsinskikh nauk, 2013; 44-51.
54. Koblihovala E., Luksan O, Mrazova I, Ryska M, Cervenka L: *Hepatocyte transplantation attenuates the course of acute liver failure induced by thioacetamide in Lewis rats*. Physiological research, 2015; 64: 689-700.
55. Adwan H, Fuller B, Seldon C, Davidson B, Seifalian A: *Modifying three-dimensional scaffolds from novel nanocomposite materials using dissolvable porogen particles for use in liver tissue engineering*. Journal of biomaterials applications, 2013; 28: 250-61. doi: 10.1177/0885328212445404.
56. Jiang, WC, et al.: *Cryo-chemical decellularization of the whole liver for mesenchymal stem cells-based functional hepatic tissue engineering*. Biomaterials; 35: 3607-617, doi: 10.1016/j.biomaterials.2014.01.024.
57. Pan J, et al.: *In-vivo organ engineering: Perfusion of hepatocytes in a single liver lobe scaffold of living rats*. The international journal of biochemistry & cell biology, 2016; 80: 124-131, doi: 10.1016/j.biocel.2016.10.003.
58. Janani G, Nandi SK, Mandal BB: *Functional hepatocyte clusters*

- on bioactive blend silk matrices towards generating bioartificial liver constructs. *Acta biomaterialia*, 2018; 67: 167-182, doi: 10.1016/j.actbio.2017.11.053.
59. Nicolas CT, et al.: Concise review: liver regenerative medicine: from hepatocyte transplantation to bioartificial livers and bioengineered grafts. *Stem cells*, 2017; 35: 42-50, doi: 10.1002/stem.2500.
60. Mazza G, et al.: Decellularized human liver as a natural 3D-scaffold for liver bioengineering and transplantation. *Scientific reports*, 2015; 5: 13079, doi: 10.1038/srep13079.
61. Chutkerashvili K, et al.: Transplantation of liver microfragments on synthetic capsules vs hepatocytes transplantation on biodegradable polymer scaffolds. *Transplantation*, 2012; 94: 1010-1010, doi: 10.1097/00007890-201211271-01997.
62. Yang W, Xia R, Zhang Y, Zhang H, Bail: Decellularized liver scaffold for liver regeneration. *Methods in molecular biology*, 2018; 1577: 11-23, doi: 10.1007/7651\_2017\_53.
63. Shirakigawa N, Ijima H, Takei T: Decellularized liver as a practical scaffold with a vascular network template for liver tissue engineering. *Journal of bioscience and bioengineering*, 2012; 114: 546-51; doi: 10.1016/j.jbiosc.2012.05.022.
64. Hammond JS, Beckingham, IJ & Shakesheff KM: Scaffolds for liver tissue engineering. *Expert review of medical devices*, 2006; 3: 21-27. doi: 10.1586/17434440.3.1.21.
65. Rad AT, et al.: Conducting scaffolds for liver tissue engineering. *Journal of biomedical materials research. Part A*, 2014; 102: 4169-181, doi: 10.1002/jbm.a.35080.
66. Versteegen MA, et al.: Decellularization of whole human liver grafts using controlled perfusion for transplantable organ bioscaffolds. *Stem cells and development*, 2017; 26: 1304-1315, doi: 10.1089/scd.2017.0095.
67. Kordzaia, D., Chanukvadze, D. & Jangavadze, M. in *Bile Duct: Functional anatomy, disease and injury classification and surgical management* (ed Miguel Ángel Mercado) Ch. 1, 279 (Nova Sciences Publisher Inc, 2014).
68. Chanukvadze I: *Construction and interactions of paravascular connective tissue structures in the liver* PhD thesis, Tbilisi State Medical Institute, 1979.
69. Patarashvili L, Tsomaia K, Kakabadze M, Kordzaia D, Chanukvadze I: *Perivascular connective tissue sheath and portal tracts in mammals*. *Translational and Clinical Medicine-Georgian Medical Journal*, 2019; 4(1):4-7.
70. Berishvili, E, et al.: Heterotopic auxiliary liver in an isolated and vascularized segment of the small intestine in rats. *Transplantation*, 2003; 75: 1827-832, doi: 10.1097/01.TP.0000065297.56712.C1.
71. Kakabadze, Z. et al.: Correction of diabetes mellitus by transplanting minimal mass of syngeneic islets into vascularized small intestinal segment. *Am J Transplant*, 2013; 13: 2550-2557, doi: 10.1111/ajt.12412.
72. Chamuleau RA: *(Bio)artificial liver support: ready for the patient?*. *Nederlands tijdschrift voor tandheelkunde*, 2016;123: 243-47. doi: 10.5177/nvt.2016.05.16100.
73. Joseph B. et al.: Isolated small intestinal segments support auxiliary livers with maintenance of hepatic functions. *Nature medicine*, 2004; 10: 749-53, doi: 10.1038/nm1057.
74. Kakabadze Z, Gupta S, Brandhors D, Korsgren O, Berishvili E: *Long-term engraftment and function of transplanted pancreatic islets in vascularized segments of small intestine*. *Transplant international: official journal of the European Society for Organ Transplantation*, 2011; 24, 175-83, doi: 10.1111/j.1432-2277.2010.01160.x.
75. Hata T, et al: *Development of a portocaval shunt using a small intestinal segment in rats*. *Microsurgery*, 2010) 30, 302-06, doi: 10.1002/micr.20751.
76. Xu S, Wu C, Zhang G, Zhang C, Huo X: *Progress in hepatocyte status detection and its application in bioartificial liver support system*. *Sheng wu yi xue gong cheng xue za zhi = Journal of biomedical engineering = Shengwu yixue gongchengxue zazhi*, 2018; 35: 151-55, doi: 10.7507/1001-5515.201705066.
77. Choi JS, Kim JD, Yoon HS Cho YW: *Full-thickness skin wound healing using human placenta-derived extracellular matrix containing bioactive molecules*. *Tissue engineering. Part A*, 2013; 19: 329-339, doi: 10.1089/ten.TEA.2011.0738.
78. Ko, I. K. et al.: *Bioengineered transplantable porcine livers with re-endothelialized vasculature*. *Biomaterials*, 2015; 40, 72-79, doi: 10.1016/j.biomaterials.2014.11.027.
79. Kakabadze Z. et al.: *Decellularized human placenta supports hepatic tissue and allows rescue in acute liver failure*. *Hepatology*, 2018; 67:1956-969. doi: 10.1002/hep.29713.
80. Syedain Z, et al.: *Tissue engineering of acellular vascular grafts capable of somatic growth in young lambs*. *Nature communications*, 2016; 7: 12951, doi: 10.1038/ncomms12951.
81. Dugladze DI, Mgeliashvili TI, Dgebuadze NN, Karagiulian SR, Managadze LG: *Treatment of acute liver insufficiency by heterotopic liver transplantation (experimental study)*. *Khirurgiia*, 1983; 47-50.
82. Diaz A, et al.: *Temporary liver transplantation in acute liver failure*. *Arch Surg*, 1977; 112, 74-78, doi: 10.1001/archsurg.1977.01370010076015.
83. Le Compte Y, deRiberolles C, Grange D, Brunet AM, Bismuth H: *Canine intrathoracic hepatic homograft: a life-supporting procedure*. *Arch Surg*, 1974; 109:80911, doi: 10.1001/archsurg.1974.01360060075020.
84. Misra M. et al.: *The effect of ischaemia on the reticuloendothelial system of the canine liver*. *Br J Surg*, 1971; 58; 867.
85. Girdaladze AM: *Temporary heterotopic liver transplantation of a newborn to an adult recipient: Exper. Research*. PhD thesis, Tbilisi medical university, 1981.
86. Ioseliani GD, Dugladze DI, Girdaladze AM, Mgeliashvili TI, & Goderdzishvili TM: *Heterotopic transplantation of the liver of a newborn infant into an adult recipient. Experimental and anatomic study*. *Khirurgiia*, 1982; 52-56.
87. Ong SL, Gravante G, Metcalfe MS, Dennison AR: *History, ethics, advantages and limitations of experimental models for hepatic ablation*. *World J Gastroenterol*, 2013; 19: 147-54, doi: 10.3748/wjg.v19.i2.147.
88. Cru RJ Jr, Garrid, AG, Silva MR: *Early hemodynamics and metabolic changes after total abdominal evisceration for experimental multivisceral transplantation*. *Acta cirurgica brasileira*, 2009; 24;156-61.
89. Partsakhashvili, D. et al.: *Experimental liver autotransplantation with novel scheme of veno-venous bypass as a model of liver denervation*.

*tion and delymphatization.* Transplant Proc, (2013) 45, 1739-1742, doi: 10.1016/j.transproceed.2012.10.048.

90. Chkhaidze Z, et al.: *New model of veno-venous bypass for management of anhepatic phase in experimental study on dogs.* Transplant Proc, 2013; 45, 1734-1738, doi: 10.1016/j.transproceed.2012.10.049.

91. Partsakhashvili DD, Azmaiparashvili EL, Chkhaidze ZA, Khodeli NG, Tomadze GD: *Optimization of the venous return during experimental hepatectomy in rabbits.* Georgian medical news, 2009; 108-112.

92. Kordzaia D, Khodeli N, Chkhaidze Z, Inauri N, Tsomaia K, Gogiashvili L: *Morphological changes in the liver after 8 hours of preservation by machine perfusion.* Georgian medical news, 2019; (295):132-37.

READ-ONLY COPY  
PRINTING PROHIBITED